

## Stereospecific Synthesis of Conformationally Constrained $\gamma$ -Amino Acids: New Foldamer Building Blocks That Support Helical Secondary Structure

Li Guo, Yonggui Chi, Aaron M. Almeida, Ilia A. Guzei, Brian K. Parker, and Samuel H. Gellman\*

Department of Chemistry, University of Wisconsin, Madison, Wisconsin 53706

Received September 2, 2009; E-mail: gellman@chem.wisc.edu

Oligomers constructed from  $\beta$ -amino acid residues (“ $\beta$ -peptides”) or from combinations of  $\alpha$ - and  $\beta$ -amino acid residues (“ $\alpha/\beta$ -peptides”) can adopt protein-like folding patterns.<sup>1,2</sup> These conformational properties provide a basis for the ongoing development of  $\beta$ - and  $\alpha/\beta$ -peptides that display interesting functional properties.  $\beta$ -Amino acid residues can be endowed with higher intrinsic folding propensities than those of  $\alpha$  residues by use of cyclic constraints to limit backbone torsional mobility, and this capacity for residue-based rigidification has proven to be important for both the structure and function of  $\beta$ - and  $\alpha/\beta$ -peptide foldamers.<sup>3–6</sup> Analogous benefits should result from the use of constrained  $\gamma$ -amino acid residues in foldamers, but it is difficult to explore this hypothesis because only a few types of ring-containing  $\gamma$ -amino acids are known.<sup>7,8</sup> The few cyclic  $\gamma$  residues examined to date have been found to promote sheet secondary structure,<sup>8b,c</sup> which contrasts with the helix-favoring effects of the most common cyclic  $\beta$  residues.<sup>1,2,5,6</sup>

Here we report a new synthetic approach that provides  $\gamma$ -amino acids containing a cyclohexyl constraint on the  $C_{\beta}$ – $C_{\gamma}$  bond and a variable side chain at  $C_{\alpha}$ . All three stereocenters of the  $\gamma$ -amino acid skeleton are generated from achiral precursors in a single process with high diastereo- and enantioselectivity. We show that the new type of  $\gamma$ -amino acid residue supports helix formation by an  $\alpha/\gamma$ -peptide backbone.

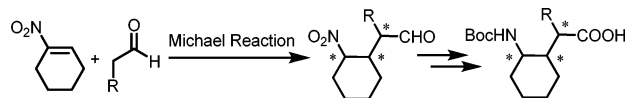
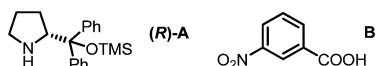


Figure 1

Figure 1 shows our synthetic approach, the key step of which is the pyrrolidine-catalyzed Michael addition of an aldehyde to 1-nitrocyclohexene. Chiral pyrrolidines have been shown to catalyze the Michael addition of aldehydes to nitroalkenes with high stereoselectivity.<sup>9,10</sup> Most precedents involve  $\beta$ -aryl nitroalkenes, such as  $\beta$ -nitrostyrene, which lead to  $\gamma^{2,3}$ -amino acids.<sup>10a–d</sup> We have reported that Michael addition of aldehydes to nitroethylene provides access to  $\gamma^2$ -amino acids.<sup>10e</sup> Use of pyrrolidine (*S*)-**A** along with acidic cocatalyst **B** proved to be optimal in terms of efficiency and enantioselectivity. Wennemers and co-workers<sup>10f</sup> concurrently devised an effective tripeptide catalyst for nitroethylene additions. In complementary work, List and co-workers<sup>10g</sup> and Hayashi et al.<sup>10h</sup> found that (*S*)-**A** catalyzes highly enantioselective Michael additions of acetaldehyde to  $\beta$ -substituted nitroalkenes, providing  $\gamma^3$ -amino acids.



Our attention was drawn to 1-nitrocyclohexene as a Michael acceptor because the adducts could be easily converted to novel

cyclically constrained  $\gamma$ -amino acid residues. Reaction of *n*-butanal and 1-nitrocyclohexene (2:1 molar ratio) in the presence of 20 mol % **A** in toluene provided the Michael adduct in only 25% yield after 24 h, and the two major diastereomers (**2a** and **3a**) were produced in a ratio of  $\sim$ 1:1 (Table 1). When 10 mol % **B** was employed as a cocatalyst, the Michael adduct yield rose to 44%, and **2a** was favored (6:1 dr); however, the major product was **4** resulting from aldol condensation. The Michael adduct yield was improved to 80% (7:1 dr) by using 5 equiv of *n*-butanal. Under these conditions, replacing **B** with either benzoic acid or acetic acid caused a modest decline in diastereoselectivity, and replacing **B** with trifluoroacetic acid completely inhibited the reaction. We speculate that a key role of the acidic component is to facilitate catalyst turnover, perhaps by promoting hydrolysis of an iminium intermediate. The selectivity for **2a** relative to the *trans* diastereomer **3a** may result from preferential equatorial protonation of the 2-substituted cyclohexane nitronate intermediate.<sup>11</sup>

Table 1. Cocatalyst Effects<sup>a</sup>

entry	cocatalyst (mol %)	yield <sup>b</sup> (%)	dr (2a/3a) <sup>b,c</sup>	M/4 <sup>d</sup>
1	none	25	1:1	1:1
2	<i>m</i> -NO <sub>2</sub> C <sub>6</sub> H <sub>4</sub> CO <sub>2</sub> H (10)	44	6:1	1:2.2
3 <sup>e</sup>	<i>m</i> -NO <sub>2</sub> C <sub>6</sub> H <sub>4</sub> CO <sub>2</sub> H (10)	80	7:1	1:3.2
4 <sup>e</sup>	HOAc (100)	80	5:1	1:5.0
5 <sup>e</sup>	TFA (10)	0	n.d.	n.d.
6 <sup>e</sup>	benzoic acid (10)	82	5:1	1:1.8

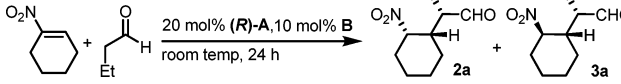
<sup>a</sup> Reactions were performed with 1.0 M 1-nitrocyclohexene using 2 equiv of aldehyde. <sup>b</sup> Determined from <sup>1</sup>H NMR analysis of the crude reaction mixture after 24 h. <sup>c</sup> See the Supporting Information for details. <sup>d</sup> **M** stands for all of the Michael adduct diastereomers observed by NMR spectroscopy. <sup>e</sup> Using 5 equiv of *n*-butanal.

Solvent choice proved to have a substantial impact on the Michael adduct yield and diastereoselectivity (Table 2). Both parameters were optimal when the reaction was conducted in CH<sub>2</sub>Cl<sub>2</sub> and catalyzed by 20 mol % **A** and 10 mol % **B**, starting with 0.5 M 1-nitrocyclohexene. These conditions led to high selectivity for the *cis* adduct **2a** (17:1 dr relative to **3a**).

Further exploration revealed that Michael additions to 1-nitrocyclohexene catalyzed by **A** are highly enantioselective and that many aldehydes are compatible with the catalytic process (Table 3). For these studies, *ee* was determined by HPLC after  $\gamma$ -nitro aldehydes had been reduced to the corresponding nitro alcohols to avoid epimerization at the  $\alpha$ -carbon. The absolute configuration of the major diastereomer generated with *n*-butanal and **A** was determined via derivatization (Scheme 1). Nitro alcohol **5** was oxidized to the corresponding nitro acid **6**, which was then coupled to L-phenylalanine methyl ester. The nitro group in the product was

hydrogenated, and the resulting amino group was protected with a Boc group. A crystal structure of this  $\alpha/\gamma$ -dipeptide revealed the *S,S,S* configuration for the  $\gamma$ -amino acid residue. The absolute configurations of other Michael adducts (Table 3) were assigned by analogy.  $\gamma$ -Nitro acid **6** could be easily converted to the Boc-protected  $\gamma$ -amino acid **7**. In terms of Michael addition scope (Table 3), it is noteworthy that aldehydes bearing a branch point adjacent to the nucleophilic carbon (such as **1d** and **1g**) are tolerated, although these examples required >2 days to produce good yields, perhaps because steric effects diminished the reactivity. The success of the aldehyde with a protected lysine-like side chain (**1i**) will facilitate the synthesis of oligomers that can be subjected to conformational analysis in aqueous solution.

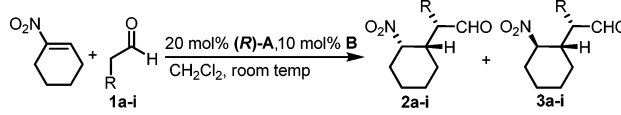
**Table 2.** Solvent Effects<sup>a</sup>



entry	solvent	concentration of 1-nitrocyclohexene (M)	yield (%) <sup>b</sup>	dr (2a/3a) <sup>b</sup>
1	toluene	1.0	80	7:1
2	hexane	1.0	80	5:1
3	DMSO	1.0	26	1:1
4	DMF	1.0	80	6:1
5	<i>i</i> -PrOH	1.0	55	4:1
6	CHCl <sub>3</sub>	1.0	85	10:1
7	CH <sub>2</sub> Cl <sub>2</sub>	1.0	86	10:1
8	CHCl <sub>3</sub>	0.5	81	15:1
9	CH <sub>2</sub> Cl <sub>2</sub>	0.5	82	17:1

<sup>a</sup> Reactions were performed using 5 equiv of aldehyde (see the Supporting Information for details). <sup>b</sup> Determined from <sup>1</sup>H NMR analysis of the crude reaction mixture after 24 h.

**Table 3.** Aldehyde Variation

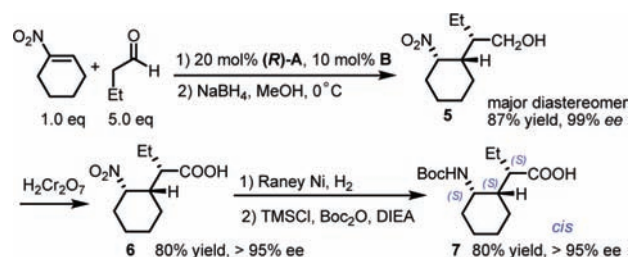


entry	product	R	time (h)	yield (%) <sup>a</sup>	dr <sup>b</sup>	ee (%) <sup>c,d</sup>
1	<b>2a</b>	Et	38	87	17:1	99
2	<b>2b</b>	Me	36	84	8:1	97
3	<b>2c</b>	<i>n</i> -Pr	40	86	16:1	99
4	<b>2d</b>	<i>i</i> -Pr	54	79	10:1	>99
5	<b>2e</b>	<i>n</i> -Bu	40	85	16:1	99
6	<b>2f</b>	<i>n</i> -Hex	42	81	15:1	>99
7	<b>2g</b>	<i>c</i> -Hex	54	70	16:1	>99
8	<b>2h</b>	CH <sub>2</sub> CH <sub>2</sub> Ph	40	75	9:1	98
9	<b>2i</b>	(CH <sub>2</sub> ) <sub>4</sub> N(Boc) <sub>2</sub>	42	73	13:1	96

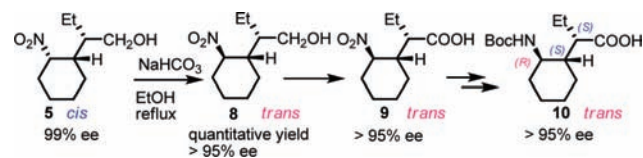
<sup>a</sup> Yield of isolated alcohol (major diastereomer) after reduction with NaBH<sub>4</sub>. <sup>b</sup> Determined from <sup>1</sup>H NMR analysis of the crude reaction mixture. <sup>c</sup> Determined by chiral HPLC analysis of the alcohols derived from **2a–i**. <sup>d</sup> Absolute configurations of **2a** and **3a** were determined by X-ray structure analysis (see the Supporting Information for details).

Overall, the results in Table 3 show that we can gain rapid access to stereochemically pure  $\gamma$ -amino acid building blocks with a *cis*-cyclohexyl constraint in the backbone and a variety of substituents adjacent to the carbonyl. The utility of the Michael addition-based approach is enhanced by the fact that the analogous *trans* diastereomers can be easily generated as well, as illustrated in Scheme 2. Thus, treating *cis*-nitro alcohol **5** with NaHCO<sub>3</sub> in ethanol at reflux quantitatively induces epimerization at the nitro-bearing carbon. Subsequent oxidation yields nitro acid **9**, which is identical to the nitro acid obtained by oxidation of **3a** (the minor product of the Michael addition, which was characterized crystallographically). Boc-protected  $\gamma$ -amino acid **10** can be readily prepared from **9**.

**Scheme 1**

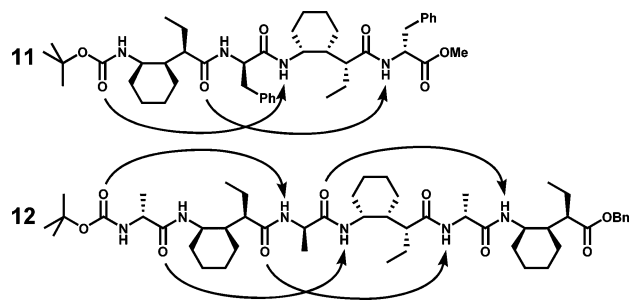


**Scheme 2**



The availability of cyclically constrained  $\gamma$ -amino acid building blocks in stereochemically pure form prompted us to begin to explore the conformational behavior of oligomers containing the corresponding subunits. Recent work suggests that oligomers constructed from  $\alpha$ - and flexible  $\gamma$ -amino acid subunits can display a variety of discrete folding patterns.<sup>12</sup> We predict that  $\alpha/\gamma$ -peptide foldamers will be conformationally stabilized by  $\gamma$ -residues with appropriate cyclic constraints.

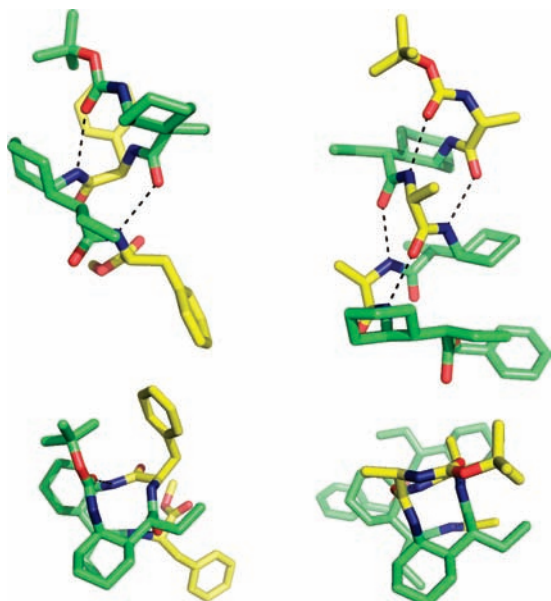
Simulations from Hofmann and co-workers<sup>13</sup> have identified a number of helical conformations that could be adopted by oligomers with a 1:1 alternation of  $\alpha$  and  $\gamma$  residues. The helix containing 12-atom-ring C=O(*i*)...H–N(*i*+3) H bonds, which may be designated the  $\alpha/\gamma$ -peptide “12-helix”, is predicted to have the *g,g* local conformation about the C <sub>$\alpha$</sub> –C <sub>$\beta$</sub>  ( $\zeta$ ) and C <sub>$\beta$</sub> –C <sub>$\gamma$</sub>  ( $\theta$ ) bonds. Fundamental principles lead one to expect that  $\gamma$  residues derived from **7** (Figure 2) will favor this local conformation. We hypothesized that the  $\alpha/\gamma$ -peptide 12-helix secondary structure would be favored by combining (*R,R,R*)-**7** [generated using (*S*)-**A**] with D- $\alpha$ -amino acid residues. This hypothesis was tested by preparation and analysis of tetramer **11** and hexamer **12**.



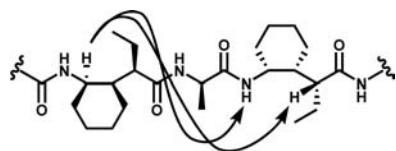
**Figure 2.** Intramolecular H-bonding patterns in the crystal structures of **11** and **12**.

The crystal structures of both **11** and **12** reveal 12-helical conformations (Figure 3); in each case, the maximum number of C=O(*i*)...H–N(*i*+3) H bonds is formed.  $\alpha/\gamma$ -Peptide **12** displayed sufficient proton resonance dispersion in CDCl<sub>3</sub> solution to enable nuclear Overhauser effect spectroscopy (NOESY) analysis. Among the unambiguous NOEs involving backbone protons, four strong NOEs were observed between protons from different  $\gamma$  residues: C <sub>$\gamma$</sub> H(2)...NH(4), C <sub>$\gamma$</sub> H(2)...C <sub>$\alpha$</sub> H(4), C <sub>$\gamma$</sub> H(4)...C <sub>$\alpha$</sub> H(6), and C <sub>$\gamma$</sub> H(4)...NH(6) (Figure 4). The C <sub>$\gamma$</sub> H(*i*)...NH(*i*+2) distances in the crystal structure of **12** are 2.5 and 2.7 Å, and the C <sub>$\gamma$</sub> H(*i*)...C <sub>$\alpha$</sub> H(*i*+2) distances are 2.4 and 2.4 Å, which suggests

that these two NOE patterns should be characteristic of the  $\alpha/\gamma$ -peptide 12-helix in solution. Balaram and co-workers<sup>12b</sup> have recently suggested that 1:1  $\alpha/\gamma$ -peptides derived from exclusively achiral amino acids can adopt the 12-helix in chloroform, but in these cases, only nearest-neighbor  $\text{NH}(i)\cdots\text{NH}(i+1)$  NOEs were observed.



**Figure 3.** Crystal structures of (left) **11** and (right) **12**: (top) views perpendicular to the helical axis; (bottom) views along the helical axis.



**Figure 4.** Characteristic NOEs patterns observed for the 1:1  $\alpha/\gamma$ -peptide hexamer **12** in  $\text{CDCl}_3$ .

We have developed a short and general route to  $\gamma$ -amino acids that feature a cyclohexyl constraint on the  $\text{C}_\beta\text{--C}_\gamma$  bond and a variety of side chains at  $\text{C}_\alpha$ . The key step is Michael addition of an aldehyde to 1-nitrocyclohexene, a process that is catalyzed by pyrrolidine **A** and strongly favors just one of the eight possible stereoisomers. A second stereoisomer is available via epimerization at  $\text{C}_\gamma$ ; the absolute configuration is controlled by the enantiomer of catalyst **A** that is employed.  $\alpha/\gamma$ -Peptides containing our constrained  $\gamma$  residues favor a specific helical conformation. We anticipate that incorporation of these new  $\gamma$  residues into other types of heterogeneous peptidic backbones will give rise to new families of foldamers and that synthetic approaches related to those described here will provide access to  $\gamma$ -amino acids with complementary constraints that further broaden the foldamer realm.

**Acknowledgment.** This research was supported by NSF (CHE-0848847). NMR spectrometers were purchased with partial support from NIH and NSF. We thank Prof. W. Seth Horne and Dr. Soo Hyuk Choi for helpful discussions; Prof. Shannon Stahl and Richard McDonald for assistance with chiral HPLC; Galina Popova, Andrew

Reidenbach, and Weicheng Zhang for help with preparation of materials; and Prof. A. J. Andre Cobb for sharing unpublished results.

**Supporting Information Available:** Experimental procedures, compound characterizations, and crystallographic data for **11** and **12** (CIF). This material is available free of charge via the Internet at <http://pubs.acs.org>.

## References

- (1) (a) Gellman, S. H. *Acc. Chem. Res.* **1998**, *31*, 173. (b) *Foldamers: Structure, Properties and Applications*; Hecht, S., Huc, I., Eds.; Wiley-VCH: Weinheim, Germany, 2007. (c) Goodman, C. M.; Choi, S.; Shandler, S.; DeGrado, W. F. *Nat. Chem. Biol.* **2007**, *3*, 252. (d) Seebach, D.; Beck, A. K.; Bierbaum, D. J. *Chem. Biodiversity* **2004**, *1*, 1111.
- (2) Horne, W. S.; Gellman, S. H. *Acc. Chem. Res.* **2008**, *41*, 1399.
- (3) (a) Huck, B. R.; Fisk, J. D.; Gellman, S. H. *Org. Lett.* **2000**, *2*, 2607. (b) De Pol, S.; Zorn, C.; Klein, C. D.; Zerbe, O.; Reiser, O. *Angew. Chem., Int. Ed.* **2004**, *43*, 511. (c) Baldauf, C.; Gunther, R.; Hofmann, H. J. *Biopolymers* **2006**, *84*, 408. (d) Sharma, G. V. M.; Nagendar, P.; Jayaprakash, P.; Krishna, P. R.; Ramakrishna, K. V. S.; Kunwar, A. C. *Angew. Chem., Int. Ed.* **2005**, *44*, 5878.
- (4) (a) Hayen, A.; Schmitt, M. A.; Ngassa, F. N.; Thomasson, K. A.; Gellman, S. H. *Angew. Chem., Int. Ed.* **2004**, *43*, 505. (b) Schmitt, M. A.; Choi, S. H.; Guzei, I. A.; Gellman, S. H. *J. Am. Chem. Soc.* **2005**, *127*, 13130. (c) Schmitt, M. A.; Choi, S. H.; Guzei, I. A.; Gellman, S. H. *J. Am. Chem. Soc.* **2006**, *128*, 4538. (d) Horne, W. S.; Price, J. L.; Keck, J. L.; Gellman, S. H. *J. Am. Chem. Soc.* **2007**, *129*, 4178. (e) Choi, S. H.; Guzei, I. A.; Gellman, S. H. *J. Am. Chem. Soc.* **2007**, *129*, 13780.
- (5) (a) Sadowsky, J. D.; Schmitt, M. A.; Lee, H.-S.; Umezawa, N.; Wang, S.; Tomita, Y.; Gellman, S. H. *J. Am. Chem. Soc.* **2005**, *127*, 11966. (b) Sadowsky, J. D.; Fairlie, W. D.; Hadley, E. B.; Lee, H. S.; Umezawa, N.; Nikolovska-Coleska, Z.; Wang, S. M.; Huang, D. C. S.; Tomita, Y.; Gellman, S. H. *J. Am. Chem. Soc.* **2007**, *129*, 139. (c) Horne, W. S.; Price, J. L.; Gellman, S. H. *Proc. Natl. Acad. Sci. U.S.A.* **2008**, *105*, 9151. (d) Horne, W. S.; Johnson, L. M.; Ketas, T. J.; Klasse, P. J.; Lu, M.; Moore, J. P.; Gellman, S. H. *Proc. Natl. Acad. Sci. U.S.A.* **2009**, *106*, 14751.
- (6) (a) Appella, D. H.; Barchi, J. J.; Durell, S. R.; Gellman, S. H. *J. Am. Chem. Soc.* **1999**, *121*, 2309. (b) LePlae, P. R.; Fisk, J. D.; Porter, E. A.; Weisblum, B.; Gellman, S. H. *J. Am. Chem. Soc.* **2002**, *124*, 6820. (c) Lee, M.; Raguse, T. L.; Schinnerl, M.; Pomerantz, W. C.; Wang, X.; Wipf, P.; Gellman, S. H. *Org. Lett.* **2007**, *9*, 1801.
- (7) (a) Hanessian, S.; Luo, X.; Schaum, R.; Michnick, S. *J. Am. Chem. Soc.* **1998**, *120*, 8569. (b) Hintermann, T.; Gademann, K.; Jaun, B.; Seebach, D. *Helv. Chim. Acta* **1998**, *81*, 983. (c) Farrera-Sinfreu, J.; Zaccaro, L.; Vidal, D.; Salvatella, X.; Giralt, E.; Pons, M.; Albericio, F.; Royo, M. A. *J. Am. Chem. Soc.* **2004**, *126*, 6048. (d) Vasudev, P. G.; Ananda, K.; Chatterjee, S.; Aravinda, S.; Shamala, N.; Balaram, P. *J. Am. Chem. Soc.* **2007**, *129*, 4039.
- (8) (a) Hagihara, M.; Anthony, N. J.; Stout, T. J.; Clardy, J.; Schreiber, S. L. *J. Am. Chem. Soc.* **1992**, *114*, 6568. (b) Woll, M. G.; Lai, J. R.; Guzei, I. A.; Taylor, S. J. C.; Smith, M. E. B.; Gellman, S. H. *J. Am. Chem. Soc.* **2001**, *123*, 11077. (c) Qureshi, M. K. N.; Smith, M. *Chem. Commun.* **2006**, 5006.
- (9) For reviews, see: (a) Berner, O. M.; Tedeschi, L.; Enders, D. *Eur. J. Org. Chem.* **2002**, 1877. (b) Santanu, M.; Yang, J. W.; Hoffmann, S.; List, B. *Chem. Rev.* **2007**, *107*, 5471.
- (10) For selected examples of organocatalytic Michael additions of aldehydes to nitroalkenes, see: (a) Betancort, J. M.; Barbas, C. F., III. *Org. Lett.* **2001**, *3*, 3737. (b) Alexakis, A.; Andrey, O. *Org. Lett.* **2002**, *4*, 3611. (c) Wang, W.; Wang, J.; Li, H. *Angew. Chem., Int. Ed.* **2005**, *44*, 1369. (d) Hayashi, Y.; Gotoh, H.; Hayashi, T.; Shoji, M. *Angew. Chem., Int. Ed.* **2005**, *44*, 4212. (e) Chi, Y.; Guo, L.; Kopf, N.; Gellman, S. H. *J. Am. Chem. Soc.* **2008**, *130*, 5608. (f) Wiesner, M.; Revell, J. D.; Tonazzi, S.; Wennemers, H. *J. Am. Chem. Soc.* **2008**, *130*, 5610. (g) Garcia-Garcia, P.; Ladepeche, A.; Halder, R.; List, B. *Angew. Chem., Int. Ed.* **2008**, *47*, 4719. (h) Hayashi, Y.; Itoh, T.; Ohkubo, M.; Ishikawa, H. *Angew. Chem., Int. Ed.* **2008**, *47*, 4722.
- (11) (a) Bordwell, F. G.; Yee, K. C. *J. Am. Chem. Soc.* **1970**, *92*, 5939. (b) Hayashi, T.; Senda, T.; Ogasawara, M. *J. Am. Chem. Soc.* **2000**, *122*, 10716. (c) List, B.; Pojarliev, P.; Martin, H. J. *Org. Lett.* **2001**, *3*, 2423.
- (12) (a) Vasudev, P. G.; Chatterjee, S.; Shamala, N.; Balaram, P. *Acc. Chem. Res.* [Online early access]. DOI: 10.1021/ar9001153. Published Online: July 2, 2009. (b) Chatterjee, S.; Vasudev, P. G.; Raghohama, S.; Ramakrishnan, C.; Shamala, N.; Balaram, P. *J. Am. Chem. Soc.* **2009**, *131*, 5956. (c) Chatterjee, S.; Vasudev, P. G.; Raghohama, S.; Shamala, N.; Balaram, P. *Biopolymers* **2008**, *90*, 759. (d) Vasudev, P. G.; Chatterjee, S.; Ananda, K.; Shamala, N.; Balaram, P. *Angew. Chem., Int. Ed.* **2008**, *47*, 6430. (e) Chatterjee, S.; Vasudev, P. G.; Ananda, K.; Raghohama, S.; Shamala, N.; Balaram, P. *J. Org. Chem.* **2008**, *73*, 6595.
- (13) Baldauf, C.; Gunther, R.; Hofmann, H. J. *J. Org. Chem.* **2006**, *71*, 2000.

JA907233Q